



PROBIOTICS IN DIARRHEA: MYTHS AND FACTS

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ABSTRACT

Diarrhea is a common health problem worldwide. Antibiotics, nosocomial infections and microorganism are the common factors responsible for diarrhea. Moreover, imbalance in intestinal microbial flora also leads to diarrhea. Probiotics is a living microorganism administered to promote the health of the host by treating or preventing infections due to strains of pathogens. Probiotics are effective against diarrhea and act by various mechanisms like colonization resistance, production of antimicrobial substances, competitive inhibition for bacterial adhesion sites, anti-secretory effect, and inhibition of toxin binding, enhancement of immune system and tropic effects of intestinal mucosa. Various randomized double blind studies indicate the effects of bacterial and yeast probiotics against various types of diarrhea. Between both types of probiotics yeast probiotics i.e. *Saccharomyces boulardii* is very useful against different types of diarrhea.

Keywords: Diarrhea; probiotics; *Saccharomyces boulardii*

INTRODUCTION

Gastrointestinal diseases are often a consequence of a myriad of factors, which disturb the bowel's complex ecosystem. Diarrhea is associated with an increased frequency of bowel movements with the production of soft or watery stool. It may be defined as the passage of more than 300 ml of liquid faeces in 24 hours. This results in fluid and electrolyte loss that may lead ultimately to death particularly in young children¹.

Antibiotics are the most common culprit of acute diarrhea due to loss of "colonization resistance" or the protective role of normal intestinal flora against pathogenic organism². A great variety of antibiotics have been implicated, but the most frequently associated with diarrhea are penicillins (especially ampicillin or amoxicillin), cephalosporins and clindamycin³⁻⁶. Over one-third of antibiotic associated diarrhea is associated with an infection by an anaerobic bacterium, *Clostridium difficile*, which also cause nosocomial (hospital acquired) outbreak⁷⁻⁹. In addition, medications and in-hospital procedures have been associated with a higher risk of diarrhea in nosocomial outbreaks^{10,11}. Host factors such as advanced age, gender and severe underlying disease conditions have been implicated in higher risk of acquiring nosocomial diarrhea^{10,12}.

Other etiologies of diarrhea are due to infections not associated with antibiotic predisposition (e.g. Toxigenic, *E.coli* and *Vibrio cholerae*, or infection with *Entamoeba histolytica*, *Giardia lamblia* or viruses). In many instances of acute diarrhea in children, hospitalized patients or HIV-infected patients, the etiological agent has not been determined. The traditional treatment for acute diarrhea often depends on whether a known etiological agent can be identified, on the severity of symptoms and the source of the infection (community or nosocomial). Electrolyte replenishment and cessation of the inciting agent (antibiotic or medication) are often all that is required for the treatment of milder forms of diarrhea. Specific therapy may be prescribed if a specific etiological agent can be detected.

Unfortunately, these steps are not always sufficient and the diarrhea may continue and become chronic, symptoms may increase in severity and spectrum or toxic mega colon or death may ensue^{3,13}.

In an effort to prevent or treat these difficult cases of diarrhea and to also re-establish the normal homeostasis of the colonic ecosystem, innovative approaches have been tried using living, biotherapeutic agents.

Physiological roles of the intestinal microflora

There are about 400 different species of flora in the intestine of each human being, and about 10 fold more bacteria than the number of human cells^{14, 15}. The dominant flora is characterized by the presence of more than 10⁸ bacteria per gram of faeces, and is anaerobic (*Bacteroides*, *Eubacterium*, *Bifidobacterium*, *Peptostreptococcus*, etc). The establishment of the neonatal, infant and adult colonic microflora is a gradual, sequential process^{16, 17}. The intestinal micro flora has many following functions as reviewed by Midtvedt¹⁸.

- Microflora offers protection against intestinal colonization with pathogenic microorganism and regulates intestinal transit.
- The intestinal wall is enlarged in the presence of intestinal microbes.
- Migrating motor complexes, production and sensitivity to peptides are dependent on the microflora.
- Deconjugation of bile acids and promotion of the enterohepatic circulation, degradation and digestion of some undigested carbohydrates, improvement of lactase tolerance, production of vitamins and growth factors for host intestinal cells.
- Intestinal flora matures and stimulates the gut immune system.

Situations of imbalance in intestinal microflora and digestive disorders

Various digestive disorders in adults and / or children may be associated with imbalance of the intestinal flora like certain types of acute and chronic diarrhea, irritable bowel syndrome, chronic inflammatory diseases of the intestines¹⁹. Antibiotic associated diarrhea (AAD) is the most typical example of a pathological situation related to imbalance of the intestinal flora, with incidence rate of 11%²⁰. The pathophysiology of AAD may involve the implantation of an enteropathogenic organism, as the result of damage to barrier flora by the antibiotic.²¹

PROBIOTICS

The Russian Metchnikov, who was awarded the Nobel Prize for medicine in 1908, for demonstrating that some bacteria could stimulate the growth of *Vibrio cholerae*, while others did inhibit its growth, first coined the theoretical concept of "probiotics" or "biotherapy"²².

A "Probiotic" or "biotherapeutic agent" is a living microorganism administered to promote the health of the host by treating or preventing infection due to strains of pathogens²³⁻²⁵. There are increasing experimental and clinical data to support probiotics use in the prevention and treatment of many gastrointestinal disorders, including inflammatory bowel disease, infectious and antibiotic related diarrhea and post surgical disorders^{26, 27}.

In an effort to reduce the use of antibiotics in the face of increasing development of antimicrobial resistant bacteria, the WHO has advocated, where possible, a policy of microbial interference therapy; the use of non-pathogens to eliminate pathogens²⁸⁻³¹.

Classification of probiotics

Globally, biotherapeutic agents can be divided into following two groups (Table 1).

Table 1: Classification of Probiotics

Bacterial Probiotics	Yeast Probiotics
<i>Lactobacillus acidophilus</i> <i>Lactobacillus casei GG</i> <i>Bifidobacterium bifidum</i> used with <i>Streptococcus thermocephalus</i> <i>Enterococcus faecium SF68</i> .	<i>Saccharomyces boulardii</i> .

Mechanism of action of probiotics

Here we cover mechanism of action of bacterial probiotics and yeast probiotics simultaneously.

Colonization resistance

Colonization resistance is the property of the normal colonic flora for protecting against colonization by pathogens, and is due to a complex interaction of various strains of colonic bacteria, which make up the microflora. A probiotics needs to inhibit the proliferation of pathogens during a period when the normal colonic microflora is disturbed²³.

Production of antimicrobial substances

Lactobacillus casei GS has been shown to produce inhibitory substances in vitro towards a broad spectrum of gram positive and gram-negative pathogens³². It also produces in vitro hydrogen peroxide, which is bactericidal³³. Yogurt, which contains *S. thermophilus* and *L. bulgarius*, has a bactericidal activity against *Clostridium difficile* in vitro^{34, 35}.

Competitive inhibition for bacterial adhesion sites

A *Lactobacillus* strain was shown to competitively inhibit adhesion of enteropathogenic *E.coli* to pig ileum and interfere with bacterial attachment to the mucosal layer of ileal conducts³⁶.

L. acidophilus strain can attach in vitro to cells resembling to enterocytes, which other strain of *L. acidophilus* do not^{37, 38}. Exposure of *Entamoeba histolytica* trophozoites to *S. boulardii*, its membrane or yeast culture supernatants decreased the number of Arophozoites able to attach to erythrocytes in vitro³⁹. *S. boulardii* also inhibited in vivo the proliferation of *C. krusei* and *C. pseudotropicalis* but had no inhibitory action on *C. tropicalis* by the same mechanism⁴⁰.

Anti-secretory effect induced by toxins

Vibrio cholerae produces a toxin, which activates adenylate cyclase of the enterocyte and stimulates cAMP production resulting in a major secretory diarrhea. *S. boulardii* inhibit cholera induced secretion in rabbit jejunum⁴¹.

Inhibition of toxins binding to intestinal receptors

Clostridium difficile is the most frequent cause of nosocomial diarrhea in adults and the pathogen causing persistent and protracted enterocolopathies^{42,43} and pseudo- membranous enterocolitis in children⁴⁴ as well as adults⁴⁵ Carthier et al found that gnotobiotic mice, who generally die rapidly after a *C. difficile* challenge, were protected after a single dose of *S. boulardii*⁴⁷. Several studies have shown that *S. boulardii* inhibited the formation of histological lesions in the cecum due to the toxins of *C. difficile* in mice and hamster⁴⁷⁻⁴⁹.

Enhancement of the immune defense system

Oral ingestion of *S. boulardii* causes significant increase in the production of secretory IgA and of the receptor for polymeric immunoglobulins in growing rat small intestine. In a study the decreased proliferation of systemically administered *Candida albicans* by treatment with *S. boulardii* presumably proceeds by same similar mode of immune stimulation⁵⁰⁻⁵².

Trophic effect on intestinal mucosa

Very high doses of spermine and spermidine given to young test animals cause a significant increase in the length and weight of the intestinal system, and accelerated adult response of the enzymes of the microvilli (lactase, sucrase, maltase and aminopeptidase) and an increase in the secretory component of the immune globulins in both the villi and the crypt cells^{53,54}. A significant increase in secretory IgA and the secretory component in rats treated with *S. boulardii*⁵² and an increase in disaccharidase activity (lactase, sucrase, maltase) in the mucosa of the small intestine has been detected in test subject^{55,56}.

Randomized double blind studies of probiotics

Here in this review we have covered randomized double blind placebo controlled clinical studies of both bacterial and yeast probiotics separately against diarrhea caused by common etiologies (Table 2 and 3).

Side effects of bacterial probiotics

• Malabsorption and metabolic acidosis

Disturbances of balance between gram-positive and gram-negative bacteria in the natural colonic flora might be inducing by the administration of gram-positive bacteria such as lactobacilli and bifido bacteria that have a stronger growth than gram-negative bacteria. This imbalance might result in metabolic D-lactate acidosis, as a consequence of the bacterial carbohydrate metabolism⁷⁵.

• Bacteraemia

Lactobacillus bacteria are not pathogenic although endocarditis, meningitis pneumonia and sepsis have been reported^{76,77}.

Arthritis and immunosuppressive effect

Injection of the cell membranes of streptococci and *Lactobacilli* can cause chronic polyarthritis. Bacterial components may possibly be transported to the joints, where they trigger off a local reaction^{78,79}.

Side effects of yeast probiotics

The safety of *S. boulardii* has been investigated in animal models and in randomized double blind studies. In mice given 5% *S. boulardii* for 70 days in their drinking water, no translocation from the gastrointestinal tract was observed. *S. boulardii* could not be detected in the organs (liver, kidney, lungs and heart) or in the mesenteric lymph glands⁸⁰. In an immune depressed animal model (prednisolone and antibiotic decontamination) *S. boulardii* could

only be detected at very low concentration in the mesenteric lymph gland⁸¹. *S. boulardii* was given to more than 40 AIDS patients without any serious side effects being reported^{74, 82, 83}. Within a period of 10

years, during which millions of *S. boulardii* treatment has been prescribed, few cases of fungaemia with *S. boulardii* have been reported⁸⁴.

Table 2: Randomized double blind placebo controlled studies of Bacterial probiotics

Sr. no	Type of bacterial probiotics	Objective	n ^o	Inhibition by probiotics (%)	Inhibition by placebo (%)	p	Ref.
01	(Living)	A*	48	70	68	N.S.	57
	<i>L.bulgaricus</i> -	B*	50	35	29	N.S.	58
	<i>L.acidophilus</i>	C*	353	01	11	N.S.	59
02	(Living)	B*	319	53	47	N.S.	60
03	<i>L. acidophilus</i>	B*	202	25.7	23.8	N.S.	61
	(Unspecified)						
04	<i>L. acidophilus</i>	D*	712	5.4	25.50	N.S	62
	(Heat Killed)						
05	<i>Lactobacillus G.G.</i>	B*	820	41	46.50	N.S	63
06	<i>L. fumentum</i>	B*	181	23.8	23.8	N.S	64
07	<i>Streptococcus</i>	E* E*	45	8.7	27.2	N.S	65
	<i>faecium</i> SF 68	F* F*	39	24 Hrs.	24 Hrs.	N.S.	66

Objective: topic of the study; A) Prevention of diarrhea in volunteers infected with ETEC; B) Prevention of traveler's diarrhea; C) Prevention of diarrhea associated with enteral feeding; D) Acute diarrhea in children; E) Prevention of AAD; F) Mean duration of diarrhea in adult with acute diarrhea; G) Prevention of diarrhea in hospitalized children.

n^o: no of patients ; p: level of significance; AAD: antibiotic associated diarrhea; ETC: enterotoxigenic E. coli

*Parameter that was statistically evaluated

Table 3: Randomized double blind placebo controlled studies with *S. boulardii*

Objective	n ^o	<i>S. boulardii</i>	Placebo	p	Ref.
Prevention of AAD*	388	4.5%	17.5%	p<0.001	67
Prevention of enteral feeding diarrhea*	40	8.7%	16.9%	p<0.001	68
Prevention of enteral feeding diarrhea*	20	1.5%	9.1%	p<0.001	69
Patients with severe burns	1231	31.8%	42.6%	p<0.002	60
C. difficile colitis*	124	26.3%	44.8%	p<0.05	70
Children's acute diarrhea*	130	15%	60 %	p<0.01	71
Adults with acute diarrhea*	92	3%	12 %	p<0.05	72
Children's chronic diarrhea*	40	30%	90 %	p<0.001	73
AIDS patients-chronic diarrhea*	35	39%	88 %	p<0.002	74

Objective: topic of the study; n^o: no of patients; p: level of significance

AAD: antibiotic associated diarrhea; ETC: enterotoxigenic *E. coli*

*Parameter that was statistically evaluated

Safety of probiotics

The safety of viable microorganism is difficult to establish using the current assessment methods; theoretically, almost any microorganism may, under certain circumstances, either cause an infection or alter its virulence due to internal or external factors. The balance between intestinal microbes, the intestinal barrier and the whole body forms a cohabitate in which the human system accepts the microbial interference and ecology in its current form. No evidence of opportunistic infection or other ill effects by probiotics has been observed nor have harmful effects been observed in controlled clinical studies with lactobacilli and bifidobacteria^{84, 85}.

Also the nonpathogenic *S. boulardii* yeast has been shown to be safe in animal models and although the yeast has been available for over 40 years and used throughout the world, side effects are uncommon⁸⁶. Nonetheless a few cases of fungemia have been reported although only 13 well documented cases of *S. boulardii* infection have been described in the literature^{87, 88}. In one study constipation and increased thirst were reported to occur⁷⁰.

In conclusion, the reported risks of biotherapeutic agents are extremely rare. Cases reported in the literature are limited to sporadic cases of transient bacteremia or fungemia⁸⁷.

CONCLUSION

In health, the gastrointestinal tract is not only an organ of digestion and absorption which is metabolically active and has specific nutrient requirements, but it has an additional function as a major barrier, protecting the body from harmful intraluminal pathogens and large antigenic molecules.

Probiotics offer a large number of theoretical advantages in the treatment of diarrhea. They offer a possible solution to the problem of the ever-increasing resistance of bacteria to antibiotics and have the advantage that they work by multiple pathophysiological mechanisms, as a result of which the development of resistance is unlikely without altering the colonization of resistance of the intestinal flora.

These probiotics can competitively prevent pathological bacterial colonization strengthen epithelial tight junctions and stimulate subsets of T-helper cell⁸⁹⁻⁹¹

S.boulardii is the only probiotics with convincing and reproducible double blind studies⁹² and statistically significant efficiency in the prevention and treatment of diarrhea. *S. boulardii* and loperamide are the only antidiarrhoeal on the positive list⁹³; the WHO regards *S. boulardii* as a possible treatment for recurrent *C. difficile* colitis⁹⁴.

Although some in vitro studies provided very interesting results, their clinical relevance in vivo would often not be confirmed, or need superphysiological concentrations of probiotics. For this reasons only the double blind placebo controlled studies are evaluated (Table No. 2 and 3).

Except *S. boulardii*, all other probiotics still need to demonstrate their effectiveness in well-designed randomized double blind studies before they can be recommended on a large scale⁹². Not a single bacterial probiotics appears clinically effective in the prevention and treatment of diarrhea in randomized double blind studies (Table 3).

If a bacterial probiotics were to be used on a large scale in the future, one would have to be certain that it was not implicated in antibiotic resistance transfer, especially if it was intrinsically resistant to particular antibiotics. Investigation must also be made to find out whether lactic acid producing bacteria are not contraindicated in indication where malabsorption may be develop which may lead to metabolic acidosis. Research must be carried out on the immunologically toxic behavior of bacterial peptidoglycan derived from the cell wall of gram positive bacteria and the possible consequences of use in diarrhea with increased permeability (e.g. salmonellosis).

The risk of probiotics getting into the blood stream is a theoretical side effect for all probiotics. *S. boulardii*, which has been widely used for years, appears to be relatively safe : only 7 cases in a period of 10 years, during which millions of treatments per year have been prescribed, have been published in well-documented clinical cases. *S. boulardii* gives promising results in AIDS patients without any serious side effects being reported. This extensive experience of use is lacking for bacterial probiotics and very few probiotics are available which describes the risk of translocation to the blood, although a considerable risk was found in immunodepressed (cancer) patients during *Bacillus cerus* therapy, and caution is urged when using lactobacilli, more particularly *L. rhamnosus*, in immunodepressed patients.

However the use of probiotics must be carefully considered when probiotics are used in patients at high risk for opportunistic infections or when the gastrointestinal tract is badly damaged. In conclusion, the health claims of bacterial or yeast probiotics require rigorous assessment and their underlying mechanism of action need to be thoroughly understood before any final statements on the clinical value of probiotics against diarrhea. Moreover there is need to develop suitable screening and selection methods for new potential strains with properties similar to or superior to the present successful probiotics against diarrhea.

REFERENCES

- Long RL, Dipiro JT. Diarrhoea and constipation, Pharmacotherapy- A pathophysiological approach. 4th ed. Dipiro JT et al. Connecticut; Appleton and long, 1999: 599-606.
- Van der Waaij D, Horstra H, Wieggersma N. Effect of β -lactam antibiotics on the resistance of the digestive tract of mice to colonization. J Infectious Diseases 1982; 146: 417-22.
- Fekety R, Shah AB. Diagnosis and treatment of *Clostridium difficile* colitis. J American Medical Association 1993; 289: 71-75.
- Kabins SA. Outbreak of clindamycin-associated colitis. Annals of Internal Medicine 1975; 83: 830-31.
- Mulligan ME, Citron D, Gabay E, Kirby B, George WL, Finegold SM. Alterations in human fecal flora, including in growth of *Clostridium difficile*, related to cefoxitin therapy. Antimicrobial Agents and Chemotherapy 1984; 26: 343-46.
- Tedesco RJ, Barton RW, Alpers DH. Clindamycin-associated colitis. Annals of Internal Medicine 1974; 81: 429-33.
- Bartlett JG. Antibiotic-associated diarrhea. Clinical infectious Diseases 1992; 15: 573-81.
- Caetano JA, Parames MT, Babo MJ, Santos A, Ferreira AB, Freitas AA, et al. Immunopharmacological effects of *Saccharomyces boulardii* in healthy human volunteers. International J Immunopharmacol 1986; 8: 245-59.
- McFarland LV, Mulligan ME, Kwok RYY, Stamm WE. Nosocomial acquisition of *Clostridium difficile* infection. New England J Med 1989; 320: 204-10.
- McFarland LV, Surawicz CM, Stamm WE. Risk factors for *Clostridium difficile* carriage and C. Difficile-associated diarrhea in a cohort of hospitalized patients. J Infectious Diseases 1990; 162: 678-84.
- Pierce PF, Wilson R, Silva J, Garagusi VF, Rifkin D, Fekety R, Nunez-Montiel O, Dowel VR, Hughes JM. Antibiotic associated pseudomembranous colitis: an epidemiological investigation of a cluster of cases. J Infectious Diseases 1982; 145: 169-74.
- Aronsson B, Molby R, Nord CE. Anti-microbial agents and *Clostridium difficile* in acute enteric disease: epidemiological data from Sweden. J Infectious Diseases 1985; 151: 476-81.
- Drapkin MS, Worthington MG, Chang TW, Razvi SA. *Clostridium difficile* colitis mimicking acute peritonitis. Archives of Surgery 1985; 120: 1321-22.
- Simon GL, Gorbach SL. Intestinal flora in health and disease. Gastroenterology 1984; 86: 174-93.
- Goldin BR, Lichtenstein AH, Gorbach SL. Nutritional and metabolic roles of intestinal bacteria. In: Modern Nutrition in Health and Disease. Shils ME, Olson JA, Shike M (editors); Philadelphia USA: Lea and Febiger; 1994: 569-80.
- Swords WE, Wu CC, Champlin FR, Buddington RK. Postnatal changes in selected bacterial groups of the pig colonic microflora. Biol Neonate 1993; 63: 191-200.
- Zhang H, Malo C, Boyle CR, Buddington RK. Diet influences development of the pig (*Sus scrofa*) intestine during the first 76 hours after birth. J Nutr 1998; 128: 1302-10.
- Midtvedt T. Microbial functional activities. In: Probiotics, Other Nutritional Factors and Intestinal Microflora. Nestle Nutrition Workshop Series Vol. 42. Hanson LA, Yolken RH (editors). Philadelphia, PA: Lippincott-Raven; 1999: 79-96.
- Hagiage M. In: La flora intestinale-de lequilibre an desequilibre. Vigol Maloine, Paris; 1994: 1-120.
- Bernet JP, Marx J, Kempf H, Giard P, Walbanm O, Lacombe A, et al. Epidemiologic des diarrhees associees lanti biotherapie orale chez Infant. In: Journi es Francophones de Pathologic Digestive; 1995.
- Buts JP. The clinical significance of *Clostridium difficile* infections in infants and children. In: Rambaud JC, editor. Updates on *Clostridium difficile*. Springer-Verlag, Paris, 1996: 29-36.
- Attwegg M. La<<EbiotherapieE>> dans la diarrhee. Der Informierte Arzt; 1992: 13.
- Elmer GW, Surawicz CM, McFarland LV. Biotherapeutic agents. A neglected modality for the treatment and prevention of selected intestinal and vaginal infections. JAMA 1996; 275: 870-76.
- Fuller R. Probiotics in human medicine. Gut 1991; 32: 489-92.
- McFarland LV, Elmer GW. Biotherapeutic agents past, present and future. Microecology Ther. 1995; 23: 46-73.
- Messing B, Crenn P, Beua P, Boutron-Ruault MC, Rambaud JC, et al. Long-term survival and pa-renteral nutrition dependence in adult patients with the short bowel syndrome. Gastroenterology 1999; 117: 1043-1050.
- Periti P, Tonelli F. Attualit \diamond bioterapeutiche degli agenti probiotici. Ruolo del *Saccharomyces boulardii*. Farm Ter 2000; XVII (1-2): 24-45.
- Fuller R, Gibson GR. Modification of the intestinal microflora using probiotics and prebiotics. Gastroenterology 1997; 32 (Suppl 222): 28-31.
- Goldin BR. Health benefits of probiotics. Br J Nutr 1998; 80 (Suppl 2): S203-S207.
- Roberfroid MB. Prebiotics and synbiotics: concepts and nutritional properties. Br J Nutr 1998; 80 (Suppl 2): S197-S202.
- Dunne C, Murphy L, Flynn S et al. Probiotics: from myth to reality. Demonstration of functionality in animal models of disease and in human clinical trails. Antonie Van Leeuwenhoek 1999; 76: 279-92.
- Silva M, Jacobus NV, Deneke C, Gorbach SL. Antimicrobial substance from a human Lactobacillus strain. Antimicrob. Agents Chemother 1987; 31: 1231-233.

33. Vandenberg PA. Lactic acid bacteria, their metabolic products and interference with microbial growth. *FEMS Microbiol. Rev.* 1993; 12: 221-38.
34. Ramare F, Nicoli J, Dabard J, et al. Trypsin-dependent production of an antimicrobial substance by a human *Peptostreptococcus* strain in gnotobiotic rats and in vitro. *Appl. Environ. Microbiol.* 1993; 59: 2876-883.
35. Kotz, CM, Peterson LR, Moody JA, Savaiano DA, Levitt MD. Effect of yoghurt on Clindamycin-induced *Clostridium difficile* colitis in hamsters. *Digest. Dis. Sci* 37, 1992; 37: 129-32.
36. Blomberg L, Henmiksson A, Conway PL. Inhibition of adhesion of *Escherichia coli* K88 to piglet ileal mucus by *Lactobacillus* spp. *Appl. Environ. Microb.* 1993; 59: 34-39.
37. Bernet MF, Brassart D, Neeser JR, Servin AL. *Lactobacillus acidophilus* LA1 binds to cultured human intestinal cell lines and inhibits cell attachment and cell invasion by enterovirulent bacteria. *Gut* 1994; 34: 483-89.
38. Coconnier MH, Francoise B, Chauve[re G, Servin AL. Adhering heat-killed human *Lactobacillus acidophilus*. Strain LB inhibits the process of pathogenicity of diarrhoeagenic bacteria in cultured human intestinal cells. *J Diarrhoeal Dis. Res.* 1993; 11: 235-42.
39. Rigothier MC, Maccario J, Gayral P. (Inhibitory activity of *Saccharomyces* yeasts on the adhesion of *Entamoeba histolytica* trophozoites to human erythrocytes in vitro. *Parasitol. Res.* 1994; 80: 10-15.
40. Ducluzeau R, Bensaada M. Effect comparé de l'administration unique ou en continu du *Saccharomyces boulardii* sur l'établissement de diverses souches de *Candida* dans le tractus digestif de souris gnotoxéniques. *Annales de Microbiologie (Paris)* 1982; 133: 491-501.
41. Vidon N, Huchet B, Rambaud JC. Influence de *Saccharomyces boulardii* sur la sécrétion jéjunale induite chez le rat par la toxine cholérique. *Gastroentérologie clinique et Biologie* 1986; 10: 13-16.
42. Delmee M, Buts JP. *Clostridium difficile*-associated diarrhoea in children. In: *Management of Digestive and Liver Disorders in infants and Children.* Textbook JP Buts and E Sokal eds, Elsevier Sciences, Amsterdam 1993: 371-79.
43. Buts JP, Corthier G, Delm[e M. *Saccharomyces boulardii* for *Clostridium difficile*-associated enteropathies in infants. *Journal of Ped. Gastroenterol. And Nutr.* 1993; 16: 419-25.
44. Buts JP, Weber AM, Roy CC, Morin CL. Pseudomembranous enterocolitis in childhood. *Gastroenterology* 1977; 73: 823-27.
45. Onderdonck AB. Antibiotic-associated pseudomembranous colitis due to toxin-producing *Clostridia*. *N. Engl. J. Med.* 1978; 298: 531-34.
46. Corthier G, Dubos F, Ducluzeau R. Prevention of *Clostridium difficile* induced mortality in gnotobiotic mice by *Saccharomyces boulardii*. *Canadian Journal of Microbiology* 1986; 32: 894-96.
47. Massot J, Sanchez O, Couchy R, Astoin J, Parodi AL. Bakteriopharma-kologische aktivitt von *Saccharomyces boulardii* bei der Clindamycin-induzierten kolitis in hamster. *Arzeimittel Forshung* 794-97.
48. Castex F, Corthier G, Jouvert S, Elmer GW, Guibal J, Lucas F, Bastide M. Prevention of experimental pseudomembranous colitis by *Saccharomyces boulardii*: topographical histology of the mucosa, bacterial counts and analysis of toxin production. In: Dougherty SJ, Hentges DL, Lysterly DM et al. (eds). *Microecology and Therapy*, Vol 19. Institute for Microbiology Herborn-Hill, 1989: 241.
49. Corthier G, Lucas R, Jouvert S, Castex F. Effect of *Saccharomyces boulardii* treatment on the activity of *Clostridium difficile* toxin in mouse digestive tract. *Toxicon* 1992; 30:1583-1589.
50. Marteau P, Rambaud JC. Potential of using lactic acid bacteria for therapy and immuno-modulation in man. *FEMS Microbiology Reviews* 1993; 12: 207-20.
51. Kaila M, Isolauri E, Saxelin M, Arvilommi H, Vesikari T. Viable versus inactivated *Lactobacillus* strain GG in acute rotavirus diarrhoea. *Archives of Disease in Childhood* 1995; 72: 51-53.
52. Buts JP, Bernasconi P, Vaerman JP, and Dive C. Stimulation of secretory IgA and secretory component of immunoglobulins in small intestine of rats treated with *Saccharomyces boulardii*. *Digestive Diseases and Sciences* 1990; 35: 251-56.
53. Buts JP, De Keyser N, Kolanowski J, Sokal E, Van Hoof F. Maturation of villus and crypt cell functions in rat small intestine role of dietary polyamines. *Digestive Diseases and Sciences* 1993; 38: 1091-1098.
54. Tutton PJM, Barkla DH. Biogenic amines as regulators of the proliferative activity of normal and neoplastic intestinal epithelial cells (review). *Anticancer Research* 1987; 7: 1-12.
55. Buts JP, Bernasconi P, Van Craynest MP, Maldague P, De Meyer R. Response of human and rat intestinal mucosa to oral administration of *Saccharomyces boulardii*. *Pediatric Research* 1986; 20: 192-96.
56. Buts JP, De KeyserN, De Raedemaeker L. *Saccharomyces boulardii* enhances rat intestinal enzyme expression by endoluminal release of polyamines. *Pediatric Research* 1994; 36: 552-527.
57. Clements ML, Levine MM, Black RE, Robins-Browne Luis A, Cisneros RM, Drusano GL, et al. *Lactobacillus* prophylaxis for diarrhea due to enterotoxigenic *Escherichia coli*. *Antimicrobial agents and chemotherapy* 1981; 20: 104-108.
58. Pozo-Olano JD, Warram JH, Gomez RG, Cavazos MG. Effect of a *Lactobacilli* preparation on traveler's diarrhea. *Gastroenterology* 1978; 74: 829-30.
59. Heimburger DC, Sockwell DG, Geels WJ. Diarrhea with enteral feeding: Prospective reappraisal of putative causes. *Nutrition* 1994; 10: 392-96.
60. Kollaritsch HH, Kemsner P, Wiedermann G, Scheiner O. Prevention of traveler's diarrhoea; comparison of different non-antibiotic preparations. *Travel Medicine International* 1989: 9-17.
61. Katelaris PH, Salam I, Farthing MJG. *Lactobacilli* to prevent traveler's diarrhea? *The New England Journal of Medicine* 1995; 333: 1360-361.
62. Bouloche J, Mouterde O, Mallet E. Traitement des diarrhées aiguës chez le nourrisson et le jeune enfant. *Annales de Pédiatrie* 1994; 41: 457-63.
63. Oksanen PJ, Salminen S, Saxelin M, Hämäläinen P, Ihantola-Vormisto A, Muurasiemi-Isoviita L, et al. Prevention of Traveller's Diarrhoea by *Lactobacillus* GG. *Annals of Medicine* 1990; 22: 53-56.
64. Wunderlich PF, Braun L, Fumagalli I, D'Apuzzo V, Heim F, Karly M, et al. Double blind report on the efficacy of lactic acid-producing enterococcus SF68 in the prevention of antibiotic-associated diarrhoea and in the treatment of acute diarrhoea. *The J of Internal Medical Research* 1989; 17: 333-38.
65. Mitra AK, Rabbani GH. A Double-Blind, Controlled trail of bioflorin (*Streptococcus faecium* SF68) in adults with acute diarrhoea due to vibrio cholerae and enterotoxigenic *Escherichia coli*. *Gastroenterology* 1990; 99: 1149-152.
66. Saavedra JM, Bauman NA, Oung I, Perman JA, Yolken RH. Feeding of *bifidobacterium bifidum* and *Streptococcus thermophilus* to infants in hospital for prevention of diarrhea and shedding of rotavirus. *Lancet* 1994; 344: 1046-1049.
67. Adam et al. Essais cliniques contrôlé en double insu de l'ultra-leveure lyophilisée (étude multicentrique par 25 medecins de 388 cas). *M.C.D.* 1976; 5: 401-406.
68. Temp[JD, Steidel AL, Bléhaut H, Hasselman M, Lutun Ph, Maurier F. Prévention par *Saccharomyces boulardii* des diarrhées de l'alimentation entérale débit continu. *La Semaine des Hopitaux de Paris* 1983; 59: 1409-412.
69. Schlotterer M, Bernasconi P, Lebreton F, Wasserman D. Intért de *Saccharomyces boulardii* dans la tolérance digestive de la nutrition entérale débit continu chez le. *Nutr. Clin. Metabol.* 1987; 1: 31-34.
70. McFarland LV, Surawicz CM, Greenberg RN, Fekety R, Elmer GW, Moyer KA, et al. A randomized placebo-controlled trail of *Saccharomyces boulardii* in combination with standard antibiotics for *clostridium difficile* disease. *The J American Medical Association* 1994; 271: 1913-918.
71. Cetina-Sauri G, Sierra Basto G. Evaluation thérapeutique de *Saccharomyces boulardii* chez des enfants souffrant de diarrhée aigu. *Annales de Pédiatrie* 1994; 41: 397-400.

72. Gechter W, Chase D, Hagenhoff G. *Saccharomyces boulardii* in acute adult diarrhea. Munch. Med. Wschr. 1990; 132: 188-92.
73. Castanada C. Effects of *Saccharomyces boulardii* in children with chronic diarrhea, especially due to giardiasis. Revista mexicana de puericultura y pediatria 1995; 2.
74. Saint-Marc T, Bléhaut H, Musial Ch, Touraine JL. Diarrhen im Zusammenhang mit AIDS Doppelblindstudie mit *Saccharomyces boulardii* Sem. Hôp. Paris 1995; 71: 735-41.
75. Coronada BE. Antibiotic-Induced D-Lactic Acidosis. Ann Intern Med 1995; 122: 839-42.
76. Susman JI, Baron EJ, Golberg SM, Kaplan MH, Pizzarello RA. Clinical manifestations and therapy of lactobacillus endocarditis: Report of a case and review of the literature. Reviews of Infectious Diseases 1986; 8: 771-76.
77. Rahman M, Chest infection caused by *Lactobacillus casei* ss rhamnosus. British Medical Journal 1982; 284: 471-72.
78. Millis JA. Do bacteria cause chronic polyarthritis. The New England Journal of Medicine 1989; 230: 245-256.
79. Steward-Tull DES. The immunological activities of bacterial peptidoglycans. Ann. Rev. Microbiol. 1980; 34: 311-40.
80. McFarland LV, Bernasconi P. *Saccharomyces boulardii*: A review of an innovative biotherapeutic agent. Microbial Ecology in Health and Disease 1993; 3: 201-5
81. Berg R, Bernasconi P, Flower D, Gautreux M. Inhibition of *Candida albicans* translocation from the gastrointestinal tract of mice by oral administration of *Saccharomyces boulardii*. The J Infectious Diseases 1993; 168: 1314-318.
82. Elmer G, Moyer K, Vega R, Surawicz C, Collier A, Hooton M, McFarland L. Pharmacokinetic studies of *Saccharomyces boulardii* in patients with HIV related chronic diarrhea in healthy volunteers. XIX International Congress on Microbial Ecology and Disease. Rome, September 1994: 18-21 (abstract book).
83. Saint-Marc T, Rossello-Prats L, Touraine JL. Efficacité de *Saccharomyces boulardii* dans le traitement des diarrhées du SIDA. Annales de Médecine Interne (Paris) 1991; 142: 64-65.
84. Adams MR, Marteau P. On the safety of lactic acid bacteria from food. Int J Food Microbiol 1995; 27: 263-64.
85. Bozzetti F, Gavazzi C, Cozzaglio L, et al. Perioperative TPN in malnourished patients with gastrointestinal cancer: a randomized clinical trial. Clin Nutr 1997; 16(2): 10.
86. McFarland LV, Bernasconi P. *Saccharomyces boulardii*: a review of an innovative biotherapeutic agent. Microbial Ecology Health Dis 1993; 6: 157-71.
87. Surawicz CM, McFarland LV. Risks of biotherapeutic agents. In: Biotherapeutic agents and infectious diseases (Elmer GW, McFarland LV, Surawicz CM, Eds), Humana Press Inc, Totowa, NJ 1999b; 12: 263-268.
88. Hennequin C, Kauffmann-Lacroix C, Jobert A, et al. Possible role of catheters in *Saccharomyces boulardii* fungemia. Eur J Clin Microbiol Infect Dis 2000; 19: 16-20.
89. Oberholzer c, Oberholzer A, Clare-Salzler M, Moklawer LL. Apoptosis in sepsis: a new target for therapeutic exploration. FASEB J 2001; 15: 879-892.
90. Rowlands BJ, Gardiner KR. Nutritional modulation of gut inflammation. Proc Nutr Society 1998; 57: 395-401.
91. Blum S, Alvarez S, Haller D, Perez P, Schiffrin EJ. Intestinal microflora and the interaction with immunocompetent cells. Atonie van Leeuwenhoek 1999; 76: 199-205.
92. Roffe C. Biotherapy for Antibiotic associated and other Diarrhea, J of Infection 1996; 32:1-10.
93. Lupke NP, Teichman A. Diarrhoea, Die positive Liste 1996.
94. WHO. *Saccharomyces boulardii*: a valuable adjunct in recurrent *Clostridium difficile* disease? WHO Drug Information 1995; 9:15-16.