

SPECTROSCOPIC CHARACTERIZATION OF DENDRIMERS

*SURYA PRAKASH GAUTAM, ARUN K. GUPTA, SHASHANK AGRAWAL, SHRUTI SUREKA

Suresh Gyan Vihar Universee, Jaipur, Rajasthan, Smriti College of Pharmaceutical Education, Dewas Naka, Indore MP 452010, India.
Email: suryagautam@ymail.com, gautamsuryaprakashgautam@gmail.com

Received: 16 Dec 2011, Revised and Accepted: 19 Jan 2012

ABSTRACT

Dendrimers acclaimed its fascinating position in the nanoworld. By virtue of its unique polymeric architecture, it exhibits precise compositional and constitutional properties. Spectroscopy is a technique that uses the interaction of energy with a sample to perform an analysis. This review is a study of the main analytical techniques used for the characterization of the chemical composition, the morphology, the shape, the homogeneity of dendrimers, synthesis, conjugation, reaction rate, determination of the molecular weight, structural defects of dendrimer, determination of polydispersity and purity of dendrimers. It includes Ultra-violet-visible (UV-vis), Infra-red (IR), Nuclear Magnetic Resonance (NMR), Mass spectrometry, Raman spectroscopy, Fluorescence spectroscopy, Atomic Force Spectroscopy, X-ray photoelectron spectroscopy (XPS), Electron Paramagnetic Resonance (EPR) Spectroscopy, and X-Ray Absorption Spectroscopy. Dendrimer characterization with the help of spectroscopic techniques is very useful and has wide application in the field of dendrimer chemistry.

Keywords: Dendrimers, Conjugation, Analytical techniques.

INTRODUCTION

Dendrimers are branched, synthetic polymers with layered architectures¹. Dendrimer is a carefully architected, highly branched and organized, polymer sphere. By virtue of its unique polymeric architecture it exhibits precise compositional and constitutional properties^{2,3}. Dendrimers are hyper branched, highly ordered 3-D structure, having definite molecular weight, size, shape, in which all the bonds are converging to a focal point^{4,5}. The size, shape, and surface properties of the polymer are used to modulate the pharmacokinetic and pharmacodynamic behavior of drugs conjugated with or encapsulated in the polymeric carrier. Recently, a class of well-defined, monodisperse, and tree-like polymers called dendrimers has attracted attention because of the flexibility they offer in terms of their size, shape, branching, length, and surface functionality^{6, 7, 8}. Characterization of polymeric materials is vital for predicting and elucidating polymer properties and morphology until recently, no single technique could completely describe the above characteristics of a polymer sample. The word 'spectroscopy' is used as a collective term for all the analytical techniques based on the interaction of light and matter. Spectrophotometry is used for both qualitative and quantitative investigations of samples. The principal objective of the work presented here is to analytically characterize and investigate dendrimers using UV-Vis spectrometry, FT-IR spectroscopy, Differential Scanning Calorimetry (DSC), NMR spectroscopy, as well as ESI Mass spectrometry, etc. Spectroscopy techniques are vital tools for analyzing dendrimer. This provides the information about the structure, reaction rate, appearance-disappearance-reappearance chemistry of characteristic peaks, conjugation etc⁹.

Spectroscopy and Spectrometry

Various spectroscopy techniques used in dendrimer characterization are enlisted as:

1. Ultra-violet-visible (UV-vis)
2. Infra-red (IR)
3. Nuclear Magnetic Resonance (NMR)
4. Mass spectrometry
5. Raman spectroscopy
6. Fluorescence spectroscopy
7. Atomic Force Spectroscopy
8. X-ray photoelectron spectroscopy (XPS)
9. Electron Paramagnetic Resonance (EPR) Spectroscopy
10. X-Ray Absorption Spectroscopy

Dendrimers UV spectroscopy

UV-Vis spectroscopy is an important analytical tool for characterization of dendrimers. UV-vis range lies between 200-

800nm. UV-Vis spectrometry provides the proof of synthesis as well as the conjugation (surface modification) on dendrimers due to characteristic absorption maximum or shift in value of lambda max (Bathochromic shift: Red shift). UV-Vis spectrometry is used to detect the functional moieties attached on dendrimer molecules. Characteristic curves in UV-Vis exhibits the specific maximum absorption peaks at specific wavelength, which is ascribed to the contribution of the conjugated moiety. This suggests the successful conjugation of surface modifiers to dendrimers¹⁰. UV-Vis is also used to determine the conjugation of dendrimer-star polymers, shifts in the peaks supports the conjugation¹¹. UV method is also used for characterization of dendrimer-Gold Nanocomposite materials¹². UV-Vis spectral studies shows reaction rate, attaching a solvatochromic probe at the core of dendrimers from G0 to G6 shows a dramatic change in the absorption maximum from G3 to G4, which is consistent with a transition from an open to a more globular shape¹³.

Dendrimers IR spectroscopy

IR spectroscopy is an analytical method used in the determination of synthesis, for determination of functional group, conjugation and drug-dendrimer interaction of dendrimer. The infrared portion of the electromagnetic spectrum is usually in between 0.8-1000µm. In determination of drug dendrimer interaction by IR spectroscopy the identification of the vibrational signature of the drug-dendrimer interactions is only possible by comparison of the interacting systems to the spectra of the dendrimers and drug. Unfortunately, the vibrational investigation and structural understanding of PAMAM dendrimers is still very limited¹⁴. Appearance-Disappearance-Reappearance of characteristic peaks provides the proof of synthesis progress. Disappearance of nitrile groups in the synthesis of PPI dendrimers, Disappearance-Reappearance of amine groups in PAMAM dendrimers generation, Pegylation of PAMAM dendrimers, the occurrence of hydrogen bonding in PPI glycine functionalized dendrimers, or the disappearance of the aldehydes during the synthesis of PMMH dendrimers reflects the synthesis and surface modifications^{15, 16, 17}.

Dendrimers NMR spectroscopy

Nuclear Magnetic Resonance (NMR) spectroscopy is valuable technique in the characterization of dendrimers. Nuclear magnetic resonance (NMR) spectroscopy permits determination of the structure and dynamics of molecules in solution. PAMAM dendrimers and complexed PAMAMs are characterized by Rotational-Echo Double Resonance (REDOR) solid-state NMR spectroscopy¹⁸. One-dimensional (1D) and two-dimensional (2D) NMR studies are used to probe the conformation of a melamine dendrimer bearing unique NMR signals from the core to the

periphery¹⁹. ³¹P NMR was utilized for phosphorus dendritic structures, their characterization and to ascertain their purity. High-resolution solution NMR spectroscopy is used to characterize the structure of Pd dendrimer-encapsulated nanoparticles (DENs). If heteroatoms are present in the dendrimer scaffold then not only ¹H-NMR and ¹³C-NMR spectroscopy but also other NMR techniques (e.g. ¹⁵N, ¹⁹F, ²⁹Si, and ³¹P) can be used for characterizing dendrimers. Characterization of dissolved dendrimers by routine (1D)-NMR spectroscopy becomes increasingly difficult with increasing generation number²⁰.

Multidimensional NMR spectroscopy ((2D)-NMR, (3D)-NMR) is also acquiring increasing importance in the characterization of dendrimers. NOESY experiments permit quantitative determinations of internuclear distances for nuclei in different parts of the dendrimer molecule. In the interpretation of (2D)-NOESY (NOESY=nuclear Overhauser enhancement spectroscopy) spectra, a knowledge of the spatial interrelationships between protons in different parts of the dendrimer scaffold can be acquired from proton-proton NOE interactions. Principal use of diffusion NMR in dendrimer chemistry is for size determination of dissolved dendrimers²¹.

Table 1: Applications of spectroscopic techniques in dendrimer characterization

| Analytical Techniques | Interpretation |
|--|---|
| Dendrimers UV spectroscopy | -Synthesis [Characteristic curves exhibits the specific maximum absorption peaks] ¹⁰ . - Conjugation(surface modification) [Shift in peak] ¹¹ . -Reaction rate ^{12, 13} . |
| Dendrimers IR spectroscopy | -Synthesis [Characteristic peaks corresponding to functional groups] ¹⁴ . -Conjugation (surface modification) [Shifts in Characteristic peaks corresponding to functional groups] ¹⁵ . -Appearance-Disappearance-Reappearance chemistry of characteristic peaks ^{16, 17} . |
| Dendrimers NMR spectroscopy: ¹ H-NMR and ¹³ C-NMR Rotational-Echo Double Resonance (REDOR) Solid-state NMR spectroscopy One-dimensional (1D) and two-dimensional (2D) NMR (2D)-NMR-techniques [e.g. (2D)-NOESY, (2D)-TOCSY (TOCSY=Total Correlation Spectroscopy) NMR Diffusions NMR spectroscopy (e.g. PGSE = Pulsed Gradient Spin Echo; STE = Stimulated Echo; DOSY = Diffusion Ordered Spectroscopy) | -Synthesis of dendrimers [Characteristic peaks in the spectra] ¹⁸ . -Conjugation chemistry [shielding deshielding effects shifts in peaks] ¹⁹ . -Hydrodynamic radii [NMR pulse-field gradient spin-echo] ¹⁹ . -Number of protons [intensity of peaks and integral value] ²⁰ . -Conformational changes [unique NMR signals from the core to the periphery] [One-dimensional (1D) and two-dimensional (2D) NMR.] ²⁰ . (i)Isomer populations observed by 1 H NMR reveal the onset of globular Structure. (ii)NOE complexity emerges with globular structure: variable temperature NOESY studies show that the peripheral groups. (iii)Variable temperature coefficients measured for NH protons suggest that solvent is largely excluded from the interior of the dendrimer ²⁰ . -Relaxation studies show that peripheral groups are more dynamic than groups at the core ²¹ . -Mobility of group [Relaxation times (T1) measurement by 1H- and 13C NMR] Since the mobility of a dendrimer segment is proportional to its T1 value ²¹ . -Encapsulation and extraction [Increase in the NMR intensity in 1D and 2D NMR spectra] ²¹ . -Determining the molecular weight ²² . -Structural defects in dendrimers ²³ . -Determination of the polydispersity ²⁴ . -Purity of dendrimers ²⁴ . -Structure ²⁵ . -Librations of terminal groups in dendrimers ²⁶ . -Interaction between PAMAM dendrimer with lipid membranes ^{27, 28} . -Binding to PAMAM dendrimer /interaction, polymer binding mode, the binding constant /complexation ²⁹ . -The size and shape of the molecules ³⁰ . -Peripherally modification ³⁰ . -Characterize the structure ³¹ . -Interaction of the different dendrimer therapeutics with a lipid bilayer, behavior of the dendrimer agents ³¹ . -Elemental composition ^{32, 33} . -Empirical formula ³⁴ . -Chemical state ³⁵ . -Thickness of one or more thin layered dendrimers ^{36, 37} . -Determining the numbers ³⁸ . -Distributions of numbers ³⁹ . -Spatial distribution of molecule ⁴⁰ . -Structural information ⁴¹ . Local geometric and electronic structures ⁴¹ . |
| Mass spectroscopy: MALDI-TOF-MS ESI-MS | -Determining the molecular weight ²² . -Structural defects in dendrimers ²³ . -Determination of the polydispersity ²⁴ . -Purity of dendrimers ²⁴ . -Structure ²⁵ . |
| Raman Spectroscopy | -Librations of terminal groups in dendrimers ²⁶ . -Interaction between PAMAM dendrimer with lipid membranes ^{27, 28} . -Binding to PAMAM dendrimer /interaction, polymer binding mode, the binding constant /complexation ²⁹ . -The size and shape of the molecules ³⁰ . -Peripherally modification ³⁰ . -Characterize the structure ³¹ . |
| Fluorescence Spectroscopy | -Interaction between PAMAM dendrimer with lipid membranes ^{27, 28} . -Binding to PAMAM dendrimer /interaction, polymer binding mode, the binding constant /complexation ²⁹ . -The size and shape of the molecules ³⁰ . -Peripherally modification ³⁰ . -Characterize the structure ³¹ . |
| Atomic Force Spectroscopy | -Characterize the structure ³¹ . -Interaction of the different dendrimer therapeutics with a lipid bilayer, behavior of the dendrimer agents ³¹ . -Elemental composition ^{32, 33} . -Empirical formula ³⁴ . -Chemical state ³⁵ . -Thickness of one or more thin layered dendrimers ^{36, 37} . -Determining the numbers ³⁸ . -Distributions of numbers ³⁹ . -Spatial distribution of molecule ⁴⁰ . -Structural information ⁴¹ . Local geometric and electronic structures ⁴¹ . |
| X-ray Photoelectron Spectroscopy | -Elemental composition ^{32, 33} . -Empirical formula ³⁴ . -Chemical state ³⁵ . -Thickness of one or more thin layered dendrimers ^{36, 37} . -Determining the numbers ³⁸ . -Distributions of numbers ³⁹ . -Spatial distribution of molecule ⁴⁰ . -Structural information ⁴¹ . Local geometric and electronic structures ⁴¹ . |
| Electron Paramagnetic Resonance Spectroscopy. | -Thickness of one or more thin layered dendrimers ^{36, 37} . -Determining the numbers ³⁸ . -Distributions of numbers ³⁹ . -Spatial distribution of molecule ⁴⁰ . -Structural information ⁴¹ . Local geometric and electronic structures ⁴¹ . |
| X-ray Absorption Spectroscopy (XAS) | -Thickness of one or more thin layered dendrimers ^{36, 37} . -Determining the numbers ³⁸ . -Distributions of numbers ³⁹ . -Spatial distribution of molecule ⁴⁰ . -Structural information ⁴¹ . Local geometric and electronic structures ⁴¹ . |

Mass spectroscopy

Mass spectroscopy is an analytical technique that measures mass to charge ratio of charged particle. The powerful capabilities of Matrix

Assisted Laser Desorption/Ionization Time-of-Flight (MALDI-TOF) Mass Spectrometry is realized with the fast and accurate determination of molar masses, the sequencing of repeat units, and recognition of polymer additives and impurities^{22, 23}.

MALDI-TOF-MS and ESI-MS number among the few analytical methods suitable for detailed studies of structural defects in dendrimers on the basis of characteristic fragmentation pattern.

It is used in the determination of the polydispersity and the purity of dendrimers, which is defined as the percentage of defect-free dendritic material.

Mass spectroscopy is used in the determination of fragmentation pattern of different dendrimers. Fragmentation of different generations of poly (amidoamine) dendrimers was explored in five common MALDI matrices: 2, 5-dihydroxybenzoic acid (DHB), 4-hydroxy-3-methoxycinnamic acid (FER), α -cyano-4-hydroxycinnamic acid (ACH), 2, 4, 6-trihydroxyacetophenone (THAP), and 3-hydroxypicolinic acid (HPA). Determining the molecular weight of the higher-generation dendrimers are under way using MALDI TOF mass spectroscopy, GPC analysis and Diffusion-ordered NMR spectroscopy (DOSY NMR)²⁴.

Raman spectroscopy

Raman spectroscopy is a spectroscopic technique used to study vibrational, rotational, and other low-frequency modes in a system²⁵. Raman spectroscopy give relevant information about the degree of cyclodehydrogenation of polyphenylene dendrimers and the characterization of PPI and phosphorus dendrimers²⁶. The low frequency Raman spectra in $R(\nu)$ representation is used to investigate the librations of terminal groups in dendrimers. FT-Raman spectroscopy provides unique detailed information about the structure of the technologically relevant materials²⁷. Raman spectroscopy is used in determination of interaction of PAMAM dendrimers with lipid bilayer. Raman spectroscopy were applied to assess the thermodynamic changes caused by PAMAM G4 and G3, and to specify the exact location of these dendrimers into the DPPC lipid bilayer. This study is helpful to rationally design new liposomal drug carriers for bioactive molecules by combining dendrimeric and liposomal technologies²⁸.

Fluorescence spectroscopy

Fluorescence spectroscopy is a type of electromagnetic spectroscopy which analyzes fluorescence from a sample. Fluorescence spectroscopy provides valuable information regarding the interaction between the drug and dendrimers. Size and shape of molecules can be determined with the help of fluorescence spectroscopy^{29,30}.

Atomic force microscopy (AFM)

AFM provides a three-dimensional surface profile and better resolution. Atomic force microscopy (AFM) is a very useful technique to characterize the structure, Interaction of the different dendrimer therapeutics with a lipid bilayer and behavior of the dendrimer agents. Polyamidoamine dendrimer modified multi-walled carbon nanotubes (dMNTs) was fabricated and characterized by AFM³¹.

X-ray photoelectron spectroscopy (XPS)

X-ray photoelectron spectroscopy (XPS): XPS is also known as ESCA, an abbreviation for Electron Spectroscopy for Chemical analysis³². X-ray photoelectron spectroscopy (XPS) is a quantitative spectroscopic technique utilized to measures the elemental composition, empirical formula, chemical state, thickness of one or more thin layered dendrimers (1–8 nm) and electronic state of the elements that exist within dendritic framework³³. Specific groups of starburst macromolecules such as P = S, aromatic rings, C-O, and C = O can be identified and characterized by X-ray photoelectron spectroscopy (XPS). Synthesis, characterization of melamine-based dendrimers, NiSn Dendrimer, electro catalysis using Pt and Pd dendrimer, Immobilization of Poly (amidoamine) dendrimers can be performed^{34, 35, 36, 37}.

Electron Paramagnetic Resonance (EPR) Spectroscopy

Electron paramagnetic resonance (EPR) or electron spin resonance (ESR) spectroscopy is a technique for studying chemical species that have one or more unpaired electrons, such as organic and inorganic

free radicals or inorganic complexes possessing a transition metal ion. EPR is found useful for dendrimer characterization-specifically, for determining the numbers, distributions of numbers, and spatial distribution of molecule^{38, 39, 40}.

X-ray Absorption Spectroscopy (XAS)

X-ray absorption spectroscopy (XAS) is a widely-used technique for determining the local geometric and/or electronic structure of matter. X-ray absorption spectroscopy (XAS) is a characterization technique used to determine structural information of a compound. It is specific for finding the *local* geometric and electronic structures⁴¹.

CONCLUSION

A spectroscopic technique has vast application in the field of dendrimer characterization. UV-Vis spectrometry provides the proof of synthesis as well as the conjugation (surface modification) on dendrimers. Appearance-Disappearance-Reappearance of characteristic peaks in IR spectra provides the proof of synthesis progress. Nuclear magnetic resonance (NMR) spectroscopy permits determination of the structure and dynamics of molecules in solution. PAMAM dendrimers and complexed PAMAMs are characterized by Rotational-Echo Double Resonance (REDOR) solid-state NMR spectroscopy. MALDI-TOF-MS and ESI-MS number among the few analytical methods suitable for detailed studies of structural defects in dendrimers on the basis of characteristic fragmentation patterns. Thus combination of all spectroscopic techniques is vital tool for the characterization of dendrimers in the era of this new molecular chemistry world.

REFERENCES

- Tomalia DA, Baker H, Dewald JR, Hall M, Kallos G, Martin S, Roeck J, Ryder JS. A new class of polymers: Starburst-dendritic macromolecules. *Polym J* 1985; 17:117–132.
- Tomalia DA, Naylor AM, Goddard III WA. Starburst dendrimers: Molecular-level control of size, shape, surface chemistry, topology, and flexibility from atoms to macroscopic matter. *Angew Chem Int Edn* 1990; 29:138-175.
- Smith PB, Martin SJ, Hall MJ, Tomalia DA. A characterization of the structure and synthetic reactions of polyamidoamine "Starburst" polymers. *App Polym Anal Charac* 1987; 357-385.
- Zhang C, Tomalia DA, Fréchet JMJ, Tomalia DA. Dendrimers and other Dendritic Polymers. John Wiley & Sons 2001; 239-253.
- Peterson J, Ebber A, Allikmaa V, Lopp M. Synthesis and CZE analysis of PAMAM dendrimers with an ethylenediamine core. *Proc Estonian Acad Sci Chem* 2001; 50:156-166.
- Twyman LJ, Beezer AE, Esfand R, Hardy MJ, Mitchell JC. The synthesis of water soluble dendrimers, and their application as possible drug delivery systems. *Tetrahedron Lett* 1999; 40:1743-1746.
- Esfand R, Tomalia D. Poly (amidoamine) (PAMAM) dendrimers: from biomimicry to drug delivery and biomedical applications. *Drug Discov Today* 2001; 6: 427-436.
- Tomalia D, Baker H, Dewald J, Hall M, Kallos G, Martin S, Roeck J, Ryder J, Smith P. A new class of polymers: starburst-dendritic macromolecules. *Polym J* 1985; 17: 117-132.
- Xiangyang S, Ganser RT, Kai S, Lajos PB, Baker JR. Characterization of crystalline dendrimer-stabilized gold nanoparticles. *J. Nanotech* 2006; 17:1072.
- Xiangyang S, Istvan JM, Patri AK, Xiangdong B, Mohammad TI, Desai A, Rose TG, James RB. Molecular heterogeneity analysis of poly(amidoamine) dendrimer-based mono- and multifunctional nanodevices by capillary electrophoresis. *The Analyst* 2006; 131:374–381.
- Ronald CH, Bauer BJ, Paul SA, Franziska G, Eric A. Templating of inorganic nanoparticles by PAMAM/PEG dendrimer-star polymers. *Polymer* 2002; 43:5473–5481.
- Tulja GR, Gowri DS, Shireesha M, Satyanarayana B. Spectrophotometric method for determination of angiotensin-II receptor antagonist in bulk and pharmaceutical dosage forms. *Int J Pharmacy Pharm Sci* 2012; 4:198-202.

13. Guoping L, Yunjun L, Huimin T. PVP and G1.5 PAMAM dendrimer co-mediated synthesis of silver nanoparticles. *J Solid State Chem* 2005; 178:1038-1043.
14. Popescu MC, Filip D, Vasile C, Cruz C, Rueff JM, Marcos M, Serrano JL, Singurel Gh. Characterization by Fourier transform Infrared Spectroscopy (FT-IR) and 2D IR correlation spectroscopy of PAMAM dendrimer. *J Phys Chem B* 2006; 110:14198-14211.
15. Kolev TM, Velcheva EA, Stamboliyska BA, Spitteller M. DFT and experimental studies of the structure and vibrational spectra of curcumin. *Int J Quant Chem* 2005; 102:1069-1079.
16. Kolhe P, Misra E, Kannan RM, Kannan S, Lieh-Lai M. Drug complexation, in vitro release and cellular entry of dendrimers and hyperbranched polymers. *Int J Pharm* 2003; 259:143-160.
17. Pesak DJ, Moore JS. Columnar liquid crystals from shape persistent dendritic molecules. *Angew Chem Int Ed Engl* 1999; 36:1636-1639.
18. Banerjee D, Broeren C, Genderen M, Meijer EW, Rinaldi PL. Multicomponent Host-Guest Chemistry of Carboxylic Acid and Phosphonic Acid Based Guests with Dendritic Hosts: An NMR Study. *Macromolecules* 2004; 37:8313-8318.
19. Victoria M, Guerra J, Aldrik H, Richard M. NMR Characterization of Fourth-Generation PAMAM dendrimers in the presence and absence of palladium dendrimer-encapsulated nanoparticles. *J Am Chem Soc* 2009; 131:341-350.
20. Karlos X, Simanek E. Conformational analysis of triazine dendrimers: Using NMR spectroscopy to probe the choreography of a dendrimer's dance. *Macromolecules* 2008; 41:4108-4114.
21. Caminade A, Laurent R, Turrin C, Rebut C, Nicot B, Ouali A, Zablocka M, Majoral J. Phosphorus dendrimers as viewed by ³¹P NMR spectroscopy; synthesis and characterization. *Comp Ren Chim* 2010; 13; 1006-1027.
22. Felder T, Schalley CA, Fakhrnabavi H, Lukin O. A combined ESI- and MALDI-MS(/MS) study of peripherally persulfonated dendrimers: False negative results by MALDI-MS and analysis of defects. *Chem Eur J* 2005; 11:1-13.
23. Hummelen JC, Dongen JL, Meijer EW. Multivalency in the gas phase: The study of dendritic aggregates by mass spectrometry multivalency in the gas phase. *Chem Eur J* 1987; 3:1489- 1493.
24. Bosman AW, Janssen HM, Meijer EW. About dendrimer: Structure, physical properties and applications. *Chem Rev* 1999; 99: 1665- 1688.
25. Davis AP, Ma G, Allen HC. Surface vibrational sum frequency and raman studies of PAMAM G0, G1 and acylated PAMAM G0 dendrimers. *Anal Chim Acta* 2003; 496:117-131.
26. Furer VL, Vandyukov AE, Majoral JP, Caminade AM, Kovalenko VI. Fourier-transform infrared and raman difference spectroscopy studies of the phosphorus-containing dendrimers. *Acta Mol Biomol Spect* 2004; 60:1649-1657.
27. Sahoo S, Chakraborti CK, Behera PK, Mishra SC. FTIR and Raman spectroscopic investigations of a controlled release polymeric suspension. *Int J Pharmacy Pharm Sci* 2011; 3:335-342.
28. Gardikis K, Hatziantoniou S, Viras K, Wagner M, Demetzos C. A DSC and raman spectroscopy study on the effect of PAMAM dendrimer on DPPC model lipid membranes. *Int J Pharm* 2006; 318: 118-123.
29. Yoshimura T, Abe S, Esumi K. Characterization of quaternized poly(amidoamine) dendrimers of generation 1 with multiple octyl chains. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 2004; 251:141-144.
30. Yuan W, Jinying Y, Mi Z, Caiyuan P. Synthesis, characterization, and fluorescence of pyrene-containing eight-arm star-shaped dendrimer-like copolymer with pentaerythritol core. *J Poly Sci* 2008; 46: 2788-2798.
31. Pan B, Cui D, Xu P, Ozkan C, Feng G, Ozkan M, Huang T, Chu B, Li Q, He R, Hu G. Synthesis and characterization of polyamidoamine dendrimer-coated multi-walled carbon nanotubes and their application in gene delivery systems. *Nanotechnology* 2009; 20:125101.
32. Caroline D, Mohamed MC, Lipskier JF, Caminade AM, Majoral JP. Characterization of dendrimers by X-ray photoelectron spectroscopy. *App Spect* 1999; 53:1277-1281.
33. Manna A, Imae T, Aoi K, Okada M, Yogo T. Synthesis of dendrimer-passivated noble metal nanoparticles in a polar medium: comparison of size between silver and gold particles. *Chem Mater* 2001; 13:1674- 1681.
34. Erick J, Sergio OA, Eric EG. Synthesis, characterization, and application of melamine-based dendrimers supported on silica gel. *J Pol Sci Part A: Pol Che* 2005; 43: 168-177.
35. Gates AT, Nettleton EG, Myers VS, Crooks RM. Synthesis and characterization of NiSn dendrimer-encapsulated nanoparticles. *Langmuir* 2010; 26:12994-12999.
36. Pande S, Weir GM, Zaccheo AB, Crooks RM. Synthesis, characterization, and electrocatalysis using Pt and Pd dendrimer-encapsulated nanoparticles prepared by galvanic exchange. *New J Chem* 2011. Advanced article.
37. Böckinga T, Elicia LS, Wongb C, Jamesc M, Watsond JA, Brownd CL, Chilcottb TC, Barrowe KD, Costerb GLH. Immobilization of dendrimers on Si-C linked carboxylic acid-terminated monolayers on silicon(111). *Thin Solid Films* 2006;515:1857-1863.
38. Ottaviani MF, Cossu E, Turro NJ, Tomalia DA. Characterization of starburst dendrimers by electron paramagnetic resonance: Positively charged nitroxide radicals of variable chain length used as spin probes. *J Am Chem Soc* 1995; 117:4387-4398.
39. Walter ED, Sebby KB, Usselman RJ, Singel DJ, Cloninger MJ. Characterization of heterogeneously functionalized dendrimers by mass spectrometry and EPR spectroscopy. *J Phys Chem B* 2005; 109: 21532-8.
40. Martini G, Ciani L. Electron spin resonance spectroscopy in drug delivery. *Phys Chem* 2009; 11: 211-254.
41. Gong QX, Khorshidi N, Stierle A, Dosch H, Chen HZ, Sellon A, He Y, Dulub O, Diebold U. The 2x1 Reconstruction of the Rutile TiO₂(011) Surface: A combined density functional theory, x-ray diffraction, and scanning tunneling microscopy study. *Surf Sci* 2009; 603:138-144.